

Substituted 5,6-(Dihydropyrido[3,4-*d*]pyrimidin-7(8*H*)-yl)-methanones as P2X7 Antagonists

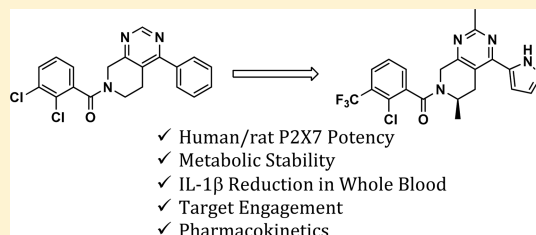
Jeannie Ziff, Dale A. Rudolph, Brice Stenne, Tatiana Koudriakova, Brian Lord, Pascal Bonaventure, Timothy W. Lovenberg, Nicholas I. Carruthers, Anindya Bhattacharya, Michael A. Letavic, and Brock T. Shireman*

Janssen Research & Development L.L.C., 3210 Merryfield Row, San Diego, California 92121, United States

Supporting Information

ABSTRACT: We describe the synthesis of a novel class of brain penetrating P2X7 antagonists with high potency at both the rat and human P2X7 receptors. Disclosed herein are druglike molecules with demonstrated target engagement of the rat P2X7 receptors after an oral dose. Specifically, compound **20** occupied the P2X7 receptors >80% over the 6 h time course as measured by an ex vivo radioligand binding experiment. In a dose–response assay, this molecule has a plasma EC₅₀ of 8 ng/mL. Overall, **20** has suitable druglike properties and pharmacokinetics in rat and dog. This molecule and others disclosed herein will serve as additional tools to elucidate the role of the P2X7 receptor in neuropsychiatric disorders.

KEYWORDS: P2X7, CNS, depression, IL-1 β , 5,6-dihydropyrido[3,4-*d*]pyrimidin-7(8*H*)-yl methanones, neuroinflammation



First cloned in 1997, the P2X7 receptor (P2X7R) is a member of the purinergic ligand-gated ion channels that are expressed on microglia and astrocytes in the central nervous system (CNS).^{1,2} Receptor activation by adenosine triphosphate (ATP) results in opening of the channel, ion flux, and the downstream release of cytokines such as IL-1 β . Expression of P2X7 on immune cells has led to extensive research efforts by pharmaceutical companies, and academia focused predominantly on its role in the pathology of disease states related to peripheral inflammatory disorders.^{3,4} Recent literature has established that P2X7 receptors in the CNS play a key role in modulating neuroinflammation via IL-1 β release through the mechanism described above.^{1,2} These findings were further corroborated by reports that IL-1 β is involved in chronic stress models.^{5,6} Furthermore, P2X7R antagonists have demonstrated efficacy in preclinical animal models of mood disorders.^{7–12} While a variety of small molecule human specific, peripherally restricted P2X7 antagonists have been disclosed and entered clinical development, optimized molecules that specifically target receptors in the CNS and have acceptable rodent pharmacology are needed.^{13,14} For example, GSK-1482160 (**1**) advanced to clinical trials for the potential treatment of neuropathic and inflammatory pain.^{15,16} A potential drawback of **1**, that would preclude its use in preclinical animal models, is the reported species potency as indicated. Compound **1** possesses 100 times lower potency for the rat P2X7 receptor versus the human. Abbott has reported on the cyanoguanidine A-804598 (**2**), which was shown to be equipotent at human and rat P2X7 receptors.¹⁷ However, significant occupancy of the P2X7 receptors in an ex vivo radioligand binding assay has only been reported following a subcutaneous dose.¹⁸

Based on the premise that a reduction of proinflammatory cytokines, such as IL-1 β , in the CNS could be an effective treatment for neuropsychiatric disorders such as depression, we set out to discover a class of small molecule brain-penetrating P2X7 receptor antagonists. In addition to potency at human P2X7 receptors, a primary goal of our program was the discovery of molecules which also possessed good potency for the rat P2X7 receptors. This would allow for the eventual evaluation of advanced compounds in preclinical models.

We have previously reported the related piperazine compounds **3** and **4**.^{11,18,19} The molecules were demonstrated to cross the blood-brain barrier and occupy the P2X7 receptors following a subcutaneous dose (sc) in an ex vivo radioligand binding experiment. In the case of **3** and **4**, sc dosing was required for extended occupancy. Our goals going forward included the discovery of a compound that engages the receptor after oral administration. An additional confounding pharmacology of **4** was its affinity for the serotonin reuptake transporter which would render any results from animal models difficult to interpret. In addition to the molecules illustrated above, we have recently reported the CNS penetrant P2X7 antagonists **5** and **6**.²⁰ Compound **5** demonstrated P2X7 target engagement after a 10 mg/kg oral dose and served as an advanced lead for further exploration. Compound **6** was shown to occupy the receptor in the same assay at 78% and 71% after a 10 mg/kg po dose at 0.5 and 2 h, respectively.

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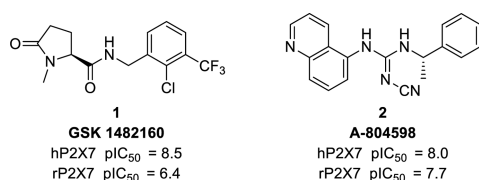


Figure 1. P2X7 antagonists GSK 1482160 and A-804598.

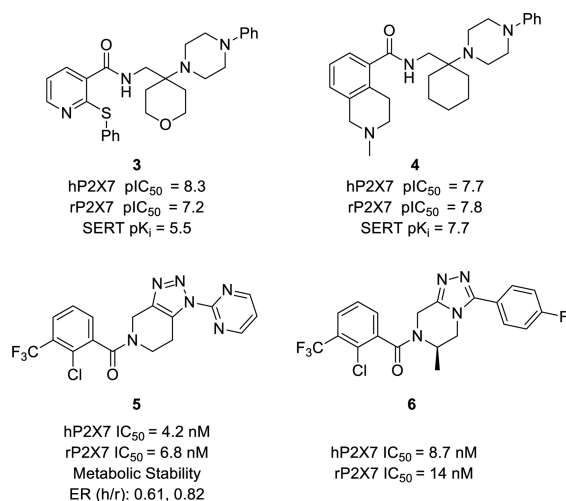


Figure 2. P2X7 antagonists from Janssen.

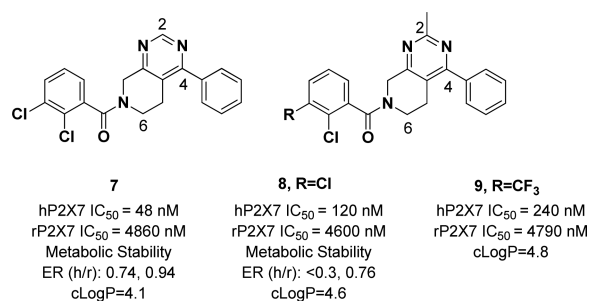


Figure 3. Starting points 7–9.

In this Letter, we report our investigations into expansion of the 5-membered ring heterocycle to pyrimidine **7**, Figure 3. In addition to in vitro data and synthesis will include selected in vivo data for this series of molecules. Compound **7** possessed moderate potency in a Ca²⁺ mobilization assay, IC₅₀ = 48 nM at the human P2X7 receptor and poor rat P2X7 potency (IC₅₀ = 4860 nM).²¹ Although **7** was shown to have a human extraction ratio (ER) of 0.74, addition of a methyl group in the 2 position of the pyrimidine resulted in compound **8** with improved stability in human microsomes (ER < 0.3). Even though this substitution resulted in a slight loss in human P2X7 potency, the substantial improvement in microsomal stability prompted a further investigation. Also illustrated in the figure is 2-chloro-3-trifluoromethyl benzamide **9**. Generally, these were used interchangeably for structure–activity relationship (SAR) purposes.

Our initial SAR began with a goal of lowering the cLog *P* by replacing the phenyl ring with a limited number of heterocycles as shown in Table 1. The *N*-2 substituted triazole **10** accomplished this goal while simultaneously improving potency at both the human and rat P2X7 receptors. Encouragingly, compound **10** also had an improved extraction ratio of <0.3

Table 1. Heterocycle Replacement at C-4

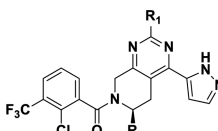
Compd	R ₁	IC ₅₀ ^{a,b} h/r (nM) ²²	hER ^c	cLogP
10		17.3/6.4	<0.3	3.2
11^d		5011/ >10,000	n.t.	3.0
12		50/170	0.71	3.4
13		8.5/1140	<0.3	3.3
14		110/21	0.52	4.5
15		46/230	0.43	3.6

^aIn vitro human and rat P2X7 IC₅₀ determined in a Ca²⁺ flux functional assay using a FLIPR instrument. ^bIC₅₀'s are the mean of at least three experiments in triplicate with a standard deviation less than 2-fold except where noted. ^cExtraction ratios are measured in a microsomal preparation. ^dRat and human P2X7 IC₅₀ from a single experiment in triplicate.

compared to the lead **7**. However, this compound was chemically unstable in solution under acidic conditions, limiting its progression. The isomeric *N*-1 isomer **11** gave a quite dramatic drop in P2X7 potency. The remaining heterocycles tested were C-linked. These included the thiazole **12**, pyrazole **13** and substituted pyrazoles **14** and **15**. Based on this data, the unsubstituted pyrazole **13** provided the best balance of human P2X7 potency (IC₅₀ = 8.5 nM) and stability in human liver microsomes (ER < 0.3). However, the rat P2X7 potency still needed improvement (IC₅₀ = 1140 nM). Moving forward, additional compounds were planned keeping the C-3 substituted pyrazole similar to **13** intact.

Further modifications to the 2-position of the pyrimidine demonstrated no real improvement in terms of human P2X7 potency as shown in Table 2. Replacement of the 2-position methyl group with a trifluoromethyl (**16**) resulted in an improvement in the rat potency, however not to a great extent. The cyclopropyl, amino, and dimethylamino (**17–19**) also offered no advantage to a methyl substituted analogue as shown. However, incorporation of a methyl group (**20**)²¹ in the 6-position, alpha to the amide nitrogen, provided the needed improvement in rat P2X7 potency, IC₅₀ = 10 nM, while maintaining human P2X7 potency, IC₅₀ = 11 nM and a moderate extraction ratio (ER = 0.56). The C-2 ethyl (**21**) and isopropyl (**22**) analogues are shown in the table as well. When comparing all properties, neither compound provided a particular advantage over the methyl substituted analog.

Table 2. Substitution at the 2 and 6 Positions



Compd	R ₁	R ₂	P2X7	
			IC ₅₀ ^{a,b} h/r (nM) ²²	hER ^c
16		H	64/240	<0.3
17		H	18/210	0.71
18		H	50/88	0.43
19		H	33/460	>0.95
20		Me	11/10	0.56
21 ^d		Me	14/13	0.78
22 ^d		Me	27/13	0.38

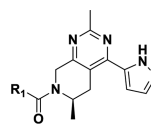
^aIn vitro human and rat P2X7 IC₅₀ determined in a Ca²⁺ flux functional assay using a FLIPR instrument. ^bIC₅₀'s are the mean of at least three experiments in triplicate with a standard deviation less than 2-fold except where noted. ^cExtraction ratios are measured in a microsomal preparation. ^dRat and human P2X7 IC₅₀ from a single experiment in triplicate.

Based on these data, the decision was made to focus on 2-methylpyrimidines with an additional methyl group on the piperidine as illustrated for compound 20. Not shown in the table is the CYP inhibition data for compound 20, which in our CYP screening assay, was determined to be a moderate CYP3A4 inhibitor (IC₅₀ of 3.3 μM).

Additional SAR was generated on the benzamide portion of the molecule as shown in Table 3. Various 2,3-disubstituted analogues were prepared to determine the effect on potency, ER and CYP 3A4 inhibition. The 2,3-dichloro analogue 24 was found to maintain potency and possessed improved microsomal stability relative to 20. However, 24 was still a CYP inhibitor (3A4 IC₅₀ = 1.9 μM). Substitutions with a 2-chloro-3-fluoro (25), 2-methyl-3-trifluoromethyl (26) or 2-fluoro-3-trifluoromethyl (27) did not provide any added benefit over 20 or 24. Interestingly, the 2-chloro-4-fluoro and 2,4-difluoro analogues 28 and 29 gave a clean CYP profile and were stable in human liver microsomes (ER < 0.3). However, this intriguing result came at the expense of P2X7 potency. Potency could be restored with the 2-fluoro-4-chloro analogue 30 while maintaining a clean CYP profile and moderate stability in human liver microsomes (ER = 0.38). The 2,4-dichloro analogue 31 was made and found to be a CYP 3A4 inhibitor with otherwise good properties with regards to P2X7 potency and microsomal stability.

Based on the balance of human and rat P2X7 potency and human microsomal stability, compounds 20 and 30 were chosen for additional profiling.²³ Selected in vitro data for 20 and 30 are shown in Table 4. Compound 20 was found to have

Table 3. Benzamide SAR



Compd	R ₁	P2X7		
		IC ₅₀ ^{a,b} h/r (nM) ²²	hER ^c	CYP 3A4 IC ₅₀ (μM)
24		18/7.9	<0.3	1.9
25		53/220	<0.3	1.8
26		17/9.2	0.65	4.2
27		18/5.9	0.41	>10 ^d
28		58/170	<0.3	>10
29		56/96	<0.3	>10
30		19/17	0.38	>10
31		11/14	<0.3	4.0

^aIn vitro human and rat P2X7 IC₅₀ determined in a Ca²⁺ flux functional assay using a FLIPR instrument. ^bIC₅₀'s are the mean of at least three experiments in triplicate with a standard deviation less than 2-fold except where noted. ^cExtraction ratios are measured in a microsomal preparation. ^d2C9 IC₅₀ = 3.4 μM

Table 4. In Vitro Data for 20 and 30

parameter	20	30
PPB h/r (% free)	6.5/5.6	21.6/15.2
BTB (% free)	3.2	7.4
CACO-2 B-A/B-A ^a	36.7/41.4	38.8/42.8
h/r (ER) ^b	0.56/0.58	0.38/0.41
P2X7 binding h/r K _i (nM) ^c	14.0/2.8	9.0/1.2
P2X7 whole blood assay IC ₅₀ (nM) ^{18d}	158	n.t.

^aP_{app} reported in units of cm/s × 10⁻⁶. ^bExtraction ratios are measured in a microsomal preparation. ^cK_i's are the mean of at least three experiments in triplicate with a standard deviation less than 2-fold. ^dHuman whole blood IC₅₀'s are a result of 2 assay runs within 2-fold.

a high free fraction in human and rat. Additionally, brain tissue binding and permeability were deemed appropriate for a CNS compound. As indicated previously, stability in liver microsomes was considered moderate for 20 and CYP 3A4 inhibition

Table 5. Rat Pharmacokinetics for 20 and 30

compd	%F (po)	CL ^c	C _{max} ^d	V _{ss} ^e
20 ^a	115	26	768	1.7
30 ^b	84	13.2	1100	1.6

^aCompound was dosed as a solution in 20% HP- β -CD at 1.0 mg/kg iv and 5.0 mg/kg po (3 animals per dose). ^bCompound was dosed as a solution in 20% HP- β -CD with 2 equiv HCl at 1.0 mg/kg iv and 5.0 mg/kg po (3 animals per dose). ^cClearance values are reported in mL/min/kg. ^dC_{max} is reported in ng/mL from the po arm. ^eV_{ss} is reported in L/kg.

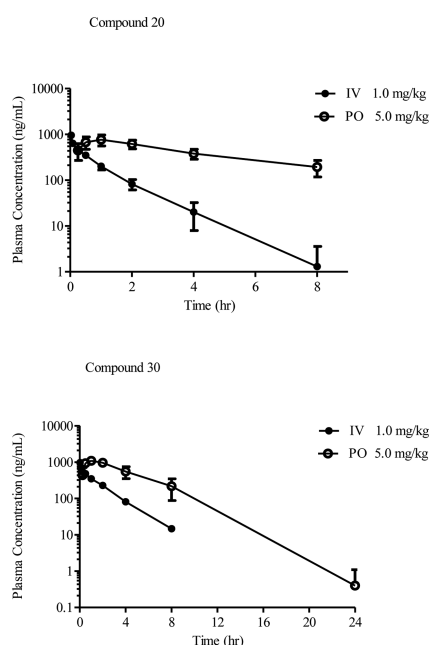


Figure 4. Rat IV and PO pharmacokinetics.

(IC₅₀ = 3.3 μ M) was viewed as a potential liability for 20. Compound 30 had the benefit from an in vitro standpoint to have less risk for CYP 3A4 inhibition (IC₅₀ > 10 μ M), a higher free fraction in plasma and brain and increased stability in microsomes. Both compounds were potent in human and rat binding assays. Compound 20 was also evaluated in a whole blood assay to determine its inhibitory effects on the release of IL-1 β . In this assay, 20 was determined to have an IC₅₀ of 158 nM. Selectivity of compounds 20 and 30 was assessed by screening against a panel of 50 ion channels and GPCRs.

Table 6. Ex Vivo Radioligand Binding Experiments

time course % occupancy/brain/plasma (ng/mL) @ time point ^a	20 ^{b,c}	30 ^{e,f}
0.5 h	81%	93%
	1877/4392	728/1860
2 h	93%	93%
	1639/3705	605/1641
6 h	83%	91%
	1427/4237	318/1089
dose response (po) ^d	20	30
ED ₅₀ (ng/mL)	0.3	0.5
B/P EC ₅₀	5/8	22/101

^aReported level of P2X7 occupancy in rat brain with 2 animals per time point. ^bCompound 20 was dosed at 30 mg/kg po in the time course. ^cCompound 30 was dosed at 10 mg/kg po in the time course. ^dReported occupancy in rat brain with 3 animals per time point. ^eP2X7 occupancy was measured at 2 h in the dose response assay. ^fP2X7 occupancy was measured at 1h in the dose response assay.

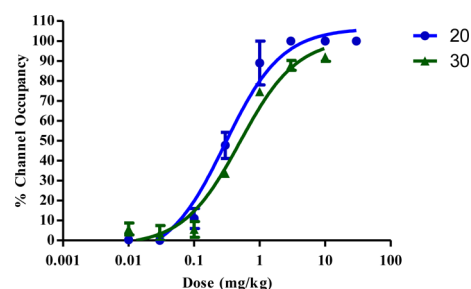


Figure 5. Ex vivo radioligand binding for 20 and 30. Dose response study after oral administration. Ex vivo occupancy was measured after compound administration at 2 h for compound 20 and 1 h for compound 30.

Table 7. Canine Pharmacokinetics for 20

compd ^a	%F	CL ^b	C _{max} ^c	V _{ss} ^d
20	35	8	571	2.3

^aCompound was dosed as a solution in 20% HP- β -CD at 0.5 mg/kg iv and 2.5 mg/kg po (3 animals per dose). ^bClearance values are reported in mL/min/kg. ^cC_{max} is reported in ng/mL from the po arm. ^dV_{ss} is reported in L/kg.

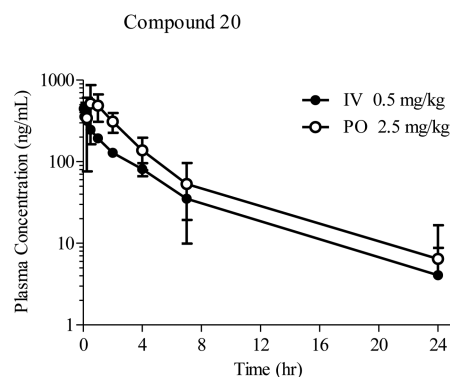


Figure 6. Canine IV and PO pharmacokinetics of 20.

Neither of the compounds showed inhibition less than 50% at 1 μ M against this panel. In addition, compound 30 was screened against the P2X1, P2X2, P2X3, P2X2/P2X3, and P2X4 ion channels and shown to have an inhibition less than 50% at 10 μ M.

Encouraged by these results, we evaluated the compounds in rat pharmacokinetic experiments, Table 5 and Figure 4. Both

compounds have good pharmacokinetics in rat following an oral dose. Gratifyingly, the clearance was moderate and predicted by in vitro rat liver microsomes. Overall oral bioavailability and exposure were consistent with levels required to occupy the receptor at low dose assuming adequate penetration into the brain.

This was confirmed in the ex vivo radioligand binding assay²⁴ as shown below in Table 6 and Figure 5. Compound 20 occupied the receptor >80% over the complete time course (6 h) of the experiment after a 10 mg/kg po dose. This was comparable to compound 30 which demonstrated >90% occupancy. Subsequent to the time course experiment, we carried out a dose response with both compounds. In this ex vivo occupancy assay, compounds 20 and 30 have an ED₅₀ of 0.3 and 0.5 mg/kg. These corresponded to plasma EC₅₀'s of 8 and 100 ng/mL for 20 and 30, respectively.

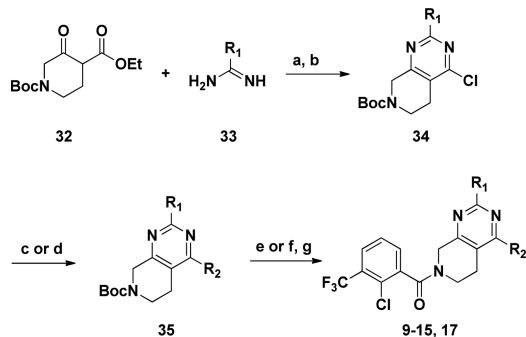
Based on the plasma EC₅₀ from the rat ex vivo radioligand binding experiment, 20 was progressed to a dog pharmacokinetic experiment as shown in Table 7 and Figure 6. Compound 20 was found to have moderate exposure (C_{max} = 571 ng/mL) after an oral dose as shown below, a low clearance (CL = 8 mL/min/kg) consistent with the measured extraction ratio (dog ER = 0.51), and an overall oral bioavailability of 35%. A human dose prediction resulted in an estimated dose of 100 mg bid in order to occupy the receptor 50% over 24 h.

METHODS

The route shown in Scheme 1 was utilized to obtain compounds 9–15, 17, and 19 by using the appropriately substituted amidine 33 at R₂ and Suzuki or Stille coupling partner. Beginning with the commercially available β-ketoester 32 and amidines 33, pyrimidine formation utilizing K₂CO₃ in MeOH followed by chlorination with PPh₃ and CCl₄ in DCE gave the 4-chloropyrimidines 34. Palladium catalyzed coupling with commercially available boronic acids or Stille reagents provided compounds 35. Deprotection and amide bond formation using the acid chloride of 2-chloro-3-trifluoromethylbenzoic acid provided the desired products 9–15, 17, and 19.

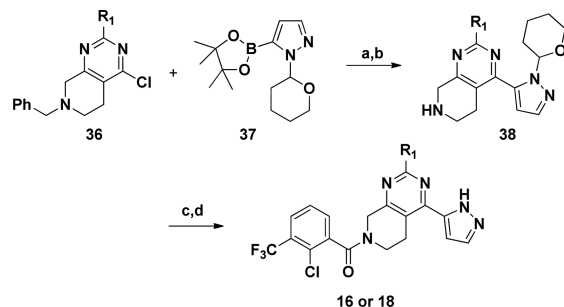
Compounds 16 and 18 were made according to the route shown in Scheme 2. Beginning with the appropriately substituted chloropyrimidine 36, Suzuki coupling with the pyrazole boronic ester 37 provided the coupling product in 67–93% yield. Deprotection of the benzyl group using 1-chloroethylchloroformate and DIEA in DCM gave compounds 38. Amide bond formation followed by removal of the THP group gave the desired molecules 16 and 18.

Scheme 1. Synthesis of Compounds 9–15, 17, and 19^a



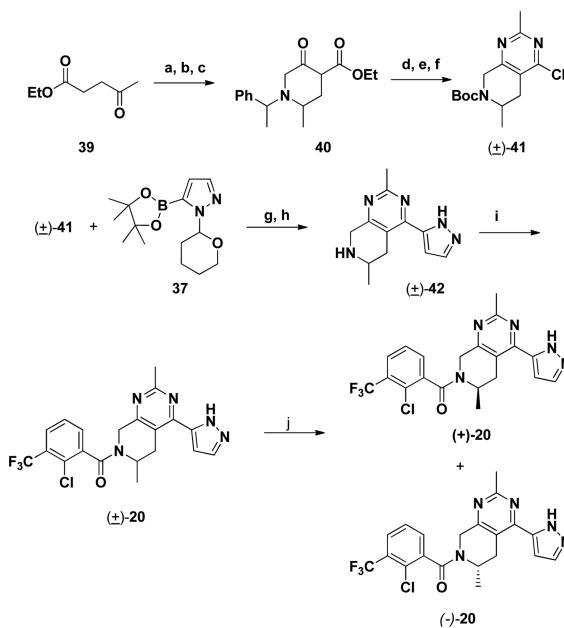
^aReagents and conditions: (a) K₂CO₃, MeOH, 70 °C; 92–99%; (b) PPh₃, CCl₄, DCE, 80 °C, 75–80%; (c) R₂BPin or R₂B(OH)₂, Pd(PPh₃)₄, 1 M Na₂CO₃ (aq), dioxane, 100 °C, 22–99%; (d) R₂SnBu₃, Pd(PPh₃)₄, dioxane, 140 °C, 15–99%; (e) TFA, DCM, quant.; (f) Et₃SiH, TFA, DCM, 85%; (g) 2-chloro-3-(trifluoromethyl)benzoic acid, HATU, DIEA, DCM, 30–99%.

Scheme 2. Synthesis of Compounds 16 and 18^a



^aReagents and conditions: (a) Pd(PPh₃)₄, 1 M Na₂CO₃ (aq), dioxane, 100 °C, 67% (R₁ = NH₂), R₂ = 93% (R₁ = CF₃); (b) 1-chloroethyl chloroformate, DIEA, DCM, MeOH, 50 °C, quant.; (c) 2-chloro-3-(trifluoromethyl)benzoic acid, HATU, DIEA, DCM, 75% (R₁ = NH₂), 37% (R₁ = CF₃); (d) Et₃SiH, TFA, DCM, 48% (R₁ = NH₂), 17% (R₁ = CF₃).

Scheme 3. Synthesis of Compound 20 and Representative Synthesis for Compounds 21–31^a



^aReagents and conditions: (a) (*R/S*)-α-methylbenzylamine, NaBH(OAc)₃, DCE, 90%; (b) ethyl glyoxylate, NaBH(OAc)₃; (c) DCE, >98%; KOtBu, PhCH₃, 63%; (d) 2-methylamidine hydrochloride, NaOEt, EtOH, 90 °C, 83%; (e) Boc₂O, ammonium formate, MeOH/THF, 10 wt % Pd/C, reflux, 82%; (f) PPh₃, CCl₄, DCE, 80 °C, 75%; (g) Pd(PPh₃)₄, 1 M Na₂CO₃ (aq), dioxane, 100 °C, 90%; (h) Et₃SiH, TFA, DCM, quantitative; (i) 2-chloro-3-trifluoromethylbenzoic acid, HATU, DIEA, DCM, 53%; (j) SFC resolution, 48% recovery for (+)-20.

Shown in Scheme 3 is the synthesis of compound 20. Beginning with ester 39, β-ketoester formation using the three-step sequence indicated provided compound 40 as a racemic mixture of diastereomers. Similar to what was described above, heterocycle formation using 2-methylamidine hydrochloride provided the intermediate pyrimidine. In this case, the benzyl group was replaced by a Boc protecting group using Pd/C and ammonium formate in the presence of Boc₂O. Chlorination of the Boc protected compound using PPh₃ and CCl₄ gave the 4-chloropyrimidine 41. Suzuki cross-coupling with the boronate ester 37 gave the desired protected pyrazole in 75% yield. Simultaneous deprotection of the THP and Boc group gave the free amine followed by amide bond formation with 2-

chloro-3-trifluoromethylbenzoic acid using HATU and DIEA in DCM gave the racemic compound (*R/S*)-**20** in 53% yield. Resolution to the individual enantiomers was carried out using SFC chromatography providing (+)-**20** and (–)-**20** in 48% and 49% yield, respectively. An analogous route was followed for compounds **21–31**.

In conclusion, we have reported a series of brain penetrating P2X7 antagonists with good potency at both the rat and human P2X7 receptors with excellent drug like properties. Compound **20** was shown to be effective in reducing IL-1 β in an in vitro whole blood assay. The compounds exhibit good oral exposure in rat and dog which leads to extended receptor occupancy and a moderate human dose prediction of 100 mg BID. Although this class of molecules did display some liabilities with regard to CYP 3A4 inhibition, these improved compounds will prove useful as additional tool molecules to delineate the role of P2X7 antagonism in neuropsychiatric disorders. Further details will be reported in due course.

■ ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge on the ACS Publications website at DOI: 10.1021/acschemneuro.5b00304.

Assay descriptions, synthetic procedures, and compound characterization (PDF)

■ AUTHOR INFORMATION

Corresponding Author

*E-mail: bshirema@its.jnj.com.

Author Contributions

J.Z., D.A.R., B.S., N.I.C., M.A.L., and B.T.S. contributed to the design and/or synthesis of the compounds in this manuscript. B.L. and P.B. were responsible for the ex vivo radioligand binding assay. A.B. and T.W.L. were responsible for the in vivo and in vitro pharmacology. T.K. was responsible for the ADME and pharmacokinetic experiments.

Notes

The authors declare no competing financial interest.

■ ACKNOWLEDGMENTS

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(23) Absolute stereochemistry was determined by analogy to ref 21.

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